

Dammarane Triterpenes from *Gardenia aubryi* VIEILL.

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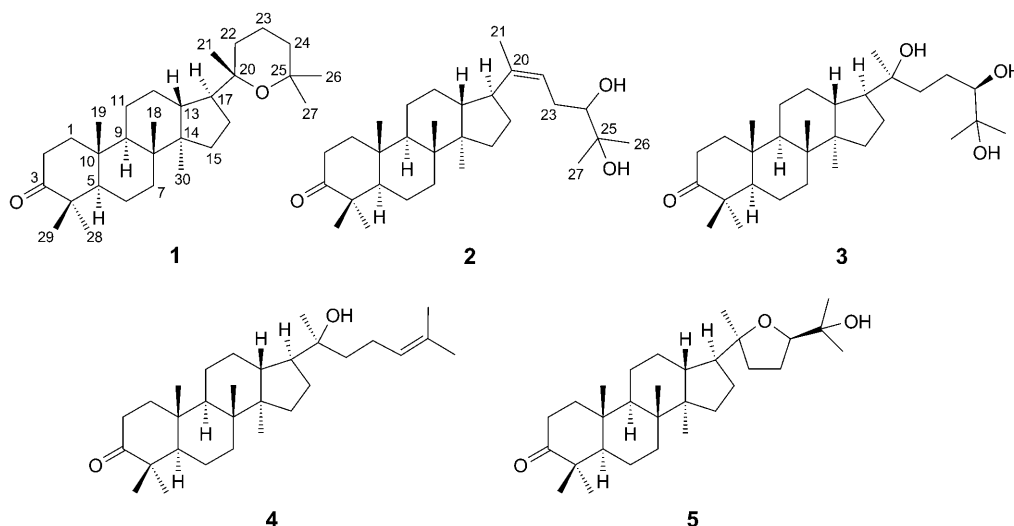
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Three new dammarane triterpenoids, gardaubryones A–C (**1–3**), were isolated from the gum collected on the aerial parts of *Gardenia aubryi* VIEILL., together with the known compounds hydroxydammarone II (**4**), ocotillone (**5**), cabraleone, and hollongdione. The structures of the novel compounds were established on the basis of mass spectrometry, NMR experiments, and chemical-correlation reactions.

Introduction. – The genus *Gardenia* (Rubiaceae) includes some two hundred species of trees and shrubs widely distributed in the warm and tropical regions of the Old World. Six endemic species occur in New Caledonia [1][2]. The leaves and flower buds of two of these, *Gardenia aubryi* VIEILL. and *Gardenia urvillei* MONTROUZ., are covered with an exudate locally used for chewing, especially by children. In a continuation of our chemical studies on secondary metabolites of New Caledonian *Gardenia* species [3], we report here the structure elucidation of three new dammarane triterpenoids, gardaubryone A–C (**1–3**), isolated from *Gardenia aubryi* gum, along with the known hydroxydammarone II (**4**), ocotillone (**5**), cabraleone, and hollongdione.

Results and Discussion. – Repeated column chromatography on silica gel of *G. aubryi* exudate resulted in the isolation of the three novel dammarane triterpenoids, gardaubryones A–C (**1–3**), together with four known dammaranes, which were identified as hydroxydammarone II (=20-hydroxy-dammar-24-en-3-one; **4** (20*S*)) [4][5], ocotillone ((=24*R*)-20,24-epoxy-25-hydroxydammaran-3-one; **5** (20*S*)) [6–8], cabraleone [6–8], and hollongdione [9][10] by comparison with literature data.

Gardaubryone A (**1**) was obtained as a colorless amorphous solid, whose empirical formula was determined as C₃₀H₅₀O₂ by HR-EI-MS. Inspection of the ¹³C-NMR spectrum revealed the presence of a C=O *s* and 29 sp³ C-atoms (six quaternary C, four CH, eleven CH₂, and eight Me). In the ¹H-NMR spectrum, the eight Me signals appeared as *ss*. A typical set of five Me resonances, correlated in the HMQC plot, at δ(C)/δ(H) 15.1/0.98, 16.1/0.91, 16.5/0.84, 21.0/1.01, and 26.7/1.13, was associated with



the Me(18), Me(19), Me(30), Me(29), and Me(28) groups, respectively, of a dammarane skeleton bearing a C=O group at C(3) (*Tables 1* and 2) [5][11][12]. Indeed, these data were closely related to those previously reported for the dammarane core present in ocotillone (**5**) [7] and in cabraleone [7], (24*S*)-20,24-epoxydammar-25(26)-en-3-one [7], 20-hydroxydammaran-3-one [5], and hydroxydammaranone II (**4**) [5]. The ^{13}C -NMR resonances of the side chain at C(17), associated with a 2,2,6-trimethyltetrahydro-2*H*-pyran moiety, were observed at $\delta(\text{C})$ 76.5 (C(20)), 26.0 (C(21)), 31.2 (C(22)), 16.4 (C(23)), 37.0 (C(24)), 70.9 (C(25)), 28.2 (C(26)), and 32.9 (C(27)). They were similar to the corresponding signals present in the spectrum of (20*S*)-panaxadiol (= (3 β ,12 β)-20,25-epoxydammarane-3,12-diol (20*S*)) [13] and strongly differed from those of (20*R*)-panaxadiol (= (3 β ,12 β)-20,25-epoxydammarane-3,12-diol) [13–15]. These data permitted to depict the structure of gardaubryone A as 20,25-epoxydammaran-3-one (20*S*) (**1**).

Gardaubyryone B (**2**) was found to possess a molecular formula $\text{C}_{30}\text{H}_{50}\text{O}_3$ by HR-EI-MS. The ^{13}C -NMR spectrum indicated the presence of a C=O, two sp^2 C-atoms (one quaternary C, one CH) corresponding to the C=C bond, and 27 sp^3 C-atoms (five quaternary C, four CH, ten CH_2 , and eight Me). The signals observed were typical for a dammaran-3-one derivative and closely related to those of **5**, except for the resonances associated with the side chain at C(17) (*Table 2*). In the ^1H -NMR spectrum, the signal of the Me(21) group at the side chain was strongly deshielded ($\delta(\text{H})$ 1.56), whereas those of the two geminal ones, Me(26) and Me(27), were only slightly deshielded. Typical ^1H , ^{13}C -HMBC cross-peaks, associated with a 6-substituted 2-methylhept-5-ene-2,3-diol moiety were noticed between Me(21) and the quaternary and tertiary sp^2 C(20) and C(22) ($\delta(\text{C})$ 141.8 and 120.5, resp.) on the one hand, and between Me(26) and Me(27) and the quaternary and tertiary oxygenated C(25) and C(24) ($\delta(\text{C})$ 72.6 and 78.2, resp.) on the other hand. The side-chain structure was confirmed by the observation of a *t* at $\delta(\text{H})$ 5.21 and a *dd* at $\delta(\text{H})$ 3.38, attributable to H–C(22) and

Table 1. ^1H -NMR Data (400 MHz, CDCl_3) of Compounds **1**–**3**. δ in ppm, J in Hz. In the case of overlapped signals or H-atoms observed as m , only the $\delta(\text{H})$ is presented. The $\delta(\text{H})$ of overlapped H-atom signals is derived from HSQC-DEPT data.

	1	2	3
$\text{CH}_2(1)$	1.93, 1.46	1.91, 1.44	1.90, 1.44
$\text{CH}_2(2)$	2.47, 2.42	2.50, 2.41	2.48, 2.43
H–C(5)	1.35	1.36	1.36
$\text{CH}_2(6)$	1.52, 1.43	1.53, 1.44	1.53, 1.45
$\text{CH}_2(7)$	1.52, 1.26	1.56, 1.30	1.54, 1.29
H–C(9)	1.40	1.38	1.42
$\text{CH}_2(11)$	1.45, 1.26	1.49, 1.18	1.48, 1.26
$\text{CH}_2(12)$	1.93, 1.26	1.84, 1.28	1.80, 1.27
H–C(13)	1.69	1.59	1.61
$\text{CH}_2(15)$	1.34, 0.98	1.47, 1.05	1.44, 1.10
$\text{CH}_2(16)$	1.64, 1.42	1.66, 1.44	1.74, 1.47
H–C(17)	1.67	2.20	1.79
Me(18)	0.98 (s)	0.99 (s)	0.97 (s)
Me(19)	0.92 (s)	0.92 (s)	0.92 (s)
Me(21)	1.11 (s)	1.56 (s)	1.15 (s)
$\text{CH}_2(22)$ or H–C(22)	1.42, 1.32	5.21 (t, $J = 7$)	1.73, 1.60
$\text{CH}_2(23)$	1.73, 1.59	2.20	1.61, 1.41
$\text{CH}_2(24)$ or H–C(24)	1.51, 1.34	3.38 (dd, $J = 8, 5.5$)	3.37 (br. d, $J = 8.5$)
Me(26)	1.14 (s)	1.17 (s)	1.22 (s)
Me(27)	1.18 (s)	1.22 (s)	1.17 (s)
Me(28)	1.01 (s)	1.02 (s)	1.02 (s)
Me(29)	1.05 (s)	1.06 (s)	1.07 (s)
Me(30)	0.84 (s)	0.86 (s)	0.87 (s)

Table 2. ^{13}C -NMR Data (50 MHz, CDCl_3) of Compounds **1**–**3**. δ in ppm.

	1	2	3		1	2	3
C(1)	39.9	39.9	39.8	C(16)	25.2	27.3	25.6
C(2)	34.1	34.1	34.1	C(17)	51.4	50.6	49.9
C(3)	218.2	218.2	218.2	C(18)	15.1	15.2	15.1
C(4)	47.4	47.4	47.4	C(19)	16.1	15.8	16.0
C(5)	55.3	55.3	55.3	C(20)	76.5	140.3	75.6
C(6)	19.7	19.6	19.6	C(21)	26.0	13.3	25.2
C(7)	34.6	34.7	34.5	C(22)	31.2	120.5	36.9
C(8)	40.3	40.4	40.2	C(23)	16.4	30.4	24.9
C(9)	50.2	50.2	50.0	C(24)	37.0	78.5	78.8
C(10)	36.9	36.9	36.8	C(25)	70.9	72.6	73.2
C(11)	22.2	21.9	22.0	C(26)	28.2	23.7	23.2
C(12)	27.6	25.8	27.4	C(27)	32.9	26.6	26.6
C(13)	42.0	44.5	42.6	C(28)	26.7	26.7	26.7
C(14)	50.0	49.2	50.2	C(29)	21.0	21.0	21.0
C(15)	31.2	31.6	31.1	C(30)	16.5	16.1	16.3

H–C(24), respectively. These data permitted to depict the structure of gardaubryone B as 24,25-dihydroxydammar-20(22)-en-3-one (**2**), in good agreement with the ^{13}C -NMR

data reported for the analogous dammar-20(22)-ene-24,25-diol moiety present in the triterpene cordialin B (= (3 β ,11 α ,20 E ,24 S)-3,19-epoxydammar-20(22)-ene-3,11,24,25-tetrol) [16]. The absolute configuration at C(24) of gardaubryone B could not be unambiguously established despite several chemical-correlation trials due to the low amount of isolated material.

It should be noted that a structure similar to that established here for gardaubryone B has been recently postulated for the triterpene ailexcelone isolated from *Ailanthus excelsa* ROXB. [17]. However, the spectral data reported for this latter compound are not consistent with the proposed structure. Specifically, the δ (C) values of the signals of the ^{13}C -NMR spectrum of ailexcelone unambiguously exclude a dammarane basic core but are in full agreement with a tirucall-7-ene (= (13 α ,14 β ,17 α ,20 S)-lanost-7-ene) structure. Consequently, ailexcelone should be regarded as one of the epimers at C(24) of 24,25-dihydroxytirucall-7-en-3-one [18]. Similarly, the structure of ailexcelol, presented as the corresponding secondary alcohol at C(3), should be revised to one of the C(24) epimers of (3 S)-tirucall-7-ene-3,24,25-triol. It should also be emphasized that (3 S ,24 S)-tirucall-7-ene-3,24,25-triol had been previously isolated from *Ailanthus excelsa* ROXB. [19].

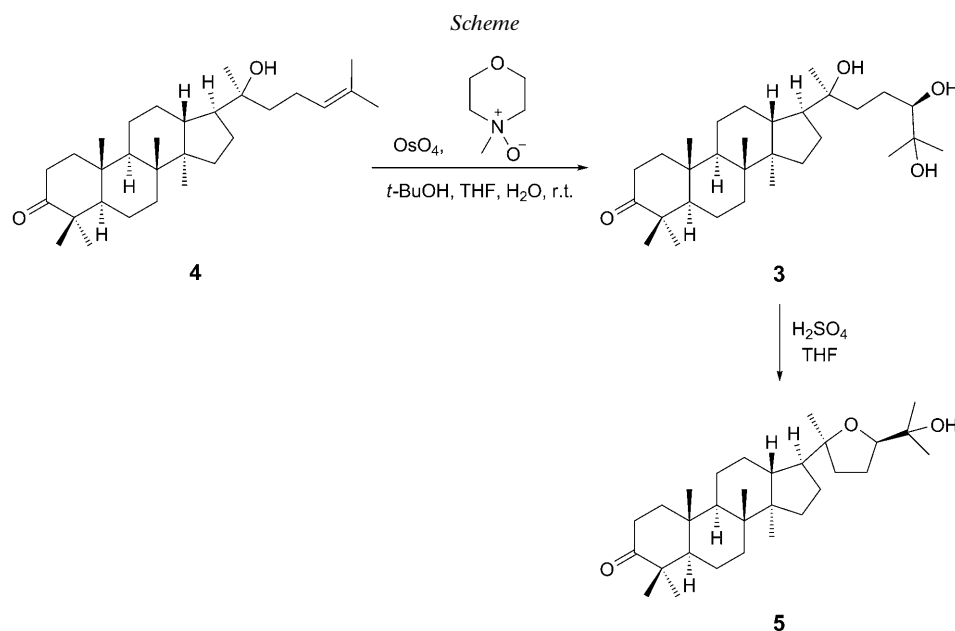
The empirical formula of gardaubryone C (**3**) was deduced as $\text{C}_{30}\text{H}_{52}\text{O}_4$ from the HR-EI-MS. The ^{13}C -NMR spectrum indicated the presence of a $\text{C}=\text{O}$ and 29 sp^3 C-atoms (six quaternary C, five CH, ten CH_2 , and eight Me). It exhibited, as those of **1** and **2**, the 22 signals of a dammaran-3-one tetracyclic skeleton. A set of additional resonances associated with a 6-substituted 2-methylheptan-2,3,6-triol moiety was noticed at δ (C) 75.6 (C(20)), 25.2 (C(21)), 36.9 (C(22)), 24.9 (C(23)), 78.8 (C(24)), 73.2 (C(25)), 23.2 (C(26)), and 26.6 (C(27)), similar to analogous signals in the spectrum of various glycosides of dammarane-3,12,20,24,25-pentol [20] and cyclo-lanostane-3,16,20,24,25-pentol [21–23]. Thus, gardaubryone C could be described as a 20,24,25-trihydroxydammaran-3-one. Catalytic OsO_4 oxidation of hydroxydammarenone II (**4**) gave a 20,24,25-trihydroxydammaran-3-one mixture of two diastereoisomers epimeric at C(24) (*Scheme*). One of these triols was identical to gardaubryone C (**3**), permitting to establish its (20 S) configuration. Upon acid treatment, gardaubryone C was converted to a mixture of the two corresponding 20,24-epoxy derivatives epimeric at C(20) [20]. One of these compounds was identical to ocotillone (**5**), giving evidence for the (24 R) configuration of gardaubryone C, the structure of which was therefore established as (24 R)-20,24,25-trihydroxydammaran-3-one (20 S).

Experimental Part

General. Liquid column chromatography (CC): silica gel 60 (SiO_2 , 40–63 μm , Merck). Optical rotations: Perkin-Elmer-341 polarimeter. IR Spectra: Perkin-Elmer-Paragon-500 instrument; $\tilde{\nu}$ in cm^{-1} . NMR Spectra: Bruker-DRX-400 and Bruker-AC-200 spectrometers; at 400 (^1H) and 50 MHz (^{13}C); δ in ppm rel. to Me_4Si as internal standard, J in Hz; 2D-NMR experiments were performed with standard Bruker micropograms. EI-MS: HP-6890 spectrometer; in m/z . HR-EI-MS: AEI-MS-902 mass spectrometer; in m/z .

Plant Material. Aerial parts of *Gardenia aubryi* were collected on July 9, 2003, in New Caledonia. A voucher sample (JWHJ-75) is kept with the Herbarium of the Center IRD of Nouméa, New Caledonia.

Extraction and Isolation. Gum (2.35 g), collected manually from fresh *Gardenia aubryi* aerial parts (1.4 kg), was dissolved in CH_2Cl_2 (35 ml) and filtered. The solvent was evaporated, and an aliquot of the



solid residue (2.10 g) was subjected to liquid vacuum chromatography (SiO_2 , stepwise gradient AcOEt /cyclohexane, 10-ml fractions). Fractions of similar composition (TLC) were pooled and further purified by CC (SiO_2 (20–45 μm), cyclohexane/ CH_2Cl_2 /MeOH (of increasing polarity): *gardaubryone A* (**1**; 28 mg), hollongdione (8 mg), ocotillone (**5**; 17 mg), cabraleone (24 mg), hydroxydammarone II (**4**; 157 mg), *gardaubryone B* (**2**; 4 mg), and *gardaubryone C* (**3**; 12 mg), in this order.

Gardaubryone A (=20,25-Epoxydammaran-3-one; **1** ((20*S*) configuration)): Amorphous solid. $[\alpha]_D^{20} = +81$ ($c=0.2$, MeOH). EI-MS: 442 (M^+). HR-EI-MS: 442.3817 (M^+ , $\text{C}_{30}\text{H}_{50}\text{O}_2^+$; calc. 442.3811).

Gardaubryone B (=24,25-Dihydroxydammar-20(22)-en-3-one; **2**): Amorphous solid. $[\alpha]_D^{20} = +84$ ($c=0.2$, MeOH). EI-MS: 458 (M^+). HR-EI-MS: 458.3758 (M^+ , $\text{C}_{30}\text{H}_{50}\text{O}_3^+$; calc. 442.3760).

Gardaubryone C ((24*R*)-20,24,25-Trihydroxydammaran-3-one; **3** ((20*S*) configuration)): Amorphous solid. $[\alpha]_D^{20} = +57$ ($c=0.2$, MeOH). EI-MS: 476 (M^+). HR-EI-MS: 442.3869 (M^+ , $\text{C}_{30}\text{H}_{52}\text{O}_4^+$; calc. 442.3866).

OsO₄ Oxidation of Hydroxydammarone II (4). To a soln. of **4** (25 mg) in $t\text{-BuOH}$ /THF/ H_2O 10:3:1 (5 ml) was added a 2.5% (w/v) soln. of OsO_4 in $t\text{-BuOH}$ (0.15 ml) and 4-methylmorpholine N -oxide (30 mg). The mixture was stirred for 48 h at 25°. A sat. aq. NaHSO_3 soln. (20 ml) was added, and the mixture was stirred for 1 h. Then, the mixture was extracted with CH_2Cl_2 (20 ml) and H_2O (20 ml), the org. layer dried (Na_2SO_4) and concentrated, and the residue submitted to flash chromatography (SiO_2 , CH_2Cl_2 /MeOH 99:1) affording **3** (8 mg), identical to the natural product isolated from *Gardenia aubryi*, and a mixture of **3** with its (24*S*) epimer which could not be separated.

Cyclization of Gardaubryone C (3). To a soln. of **3** (8 mg) in THF (5 ml) was added H_2SO_4 (0.05 ml). The mixture was stirred for 24 h at 25°. Then, the solvent was evaporated and the residue purified by prep. TLC: ocotillone (**5**; 5 mg).

REFERENCES

- [1] A. Guillaumin, 'Flore analytique et synoptique de la Nouvelle-Calédonie, Phanérogames', Office de la Recherche Scientifique Coloniale, Paris, 1948.

- [2] T. Jaffré, P. Morat, J.-M. Veillon, F. Rigault, G. Dagostini, 'Composition et caractérisation de la flore indigène de Nouvelle-Calédonie', Center IRD de Nouméa, Nouméa, 2001.
- [3] R. Grougnet, P. Magiatis, S. Mitaku, S. Loizou, P. Moutsatsou, A. Terzis, P. Cabalion, F. Tillequin, S. Michel, *J. Nat. Prod.* **2006**, 69, 1711.
- [4] J. Azakawa, R. Kasai, K. Yamazaki, O. Tanaka, *Tetrahedron* **1977**, 33, 1935.
- [5] M. Tori, R. Matsuda, M. Sono, Y. Asakawa, *Magn. Reson. Chem.* **1988**, 26, 581.
- [6] M. M. Rao, H. Meshulam, R. Zelnik, D. Lavie, *Tetrahedron* **1975**, 31, 333.
- [7] W. Aalbersberg, Y. Singh, *Phytochemistry* **1991**, 30, 921.
- [8] A. M. C. Arriaga, A. C. de Mesquita, Y. B. M. Pouliquen, R. A. de Lima, S. H. Cavalcante, M. G. de Carvalho, J. A. de Siqueira, L. V. Alegrio, R. Braz-Filho, *An. Acad. Bras. Cienc.* **2002**, 74, 415.
- [9] A. S. Gupta, S. Dev, *Tetrahedron* **1971**, 27, 823.
- [10] R. Tanaka, M. Matsuda, S. Matsunaga, *Phytochemistry* **1987**, 26, 3365.
- [11] A. Hisham, M. D. Ajitha Bai, Y. Fujimoto, N. Hara, H. Shimada, *Magn. Reson. Chem.* **1996**, 34, 146.
- [12] X.-D. Luo, S.-H. Wu, Y.-B. Ma, D.-G. Wu, *Heterocycles* **2000**, 53, 2795.
- [13] M. C. Nguyen, R. Kasai, K. Ohtani, A. Ito, T. N. Nguyen, K. Yamasaki, O. Tanaka, *Chem. Pharm. Bull.* **1994**, 42, 634.
- [14] M. Nagai, O. Tanaka, S. Shibata, *Tetrahedron Lett.* **1966**, 4797.
- [15] V. A. Denisenko, V. L. Novikov, G. V. Malinovskaya, G. B. Elyakov, *Bull. Acad. Sci. USSR, Div. Chem. Sci. (Engl. Transl.)* **1983**, 32, 2445.
- [16] V. Vande Velde, D. Lavie, R. Zelnik, A. K. Matida, S. Panizza, *J. Chem. Soc., Perkin Trans. 1* **1982**, 2697.
- [17] P. V. Srinivas, R. R. Rao, J. M. Rao, *Chem. Biodiversity* **2006**, 3, 930.
- [18] D. A. Mulholland, M. Kotsos, H. A. Mahomed, D. A. H. Taylor, *Phytochemistry* **1998**, 49, 2457.
- [19] M. H. Sherman, R. P. Borris, M. Ogura, G. A. Cordell, N. R. Farnsworth, *Phytochemistry* **1980**, 19, 1499.
- [20] S. Fujita, R. Kasai, K. Ohtani, K. Yamasaki, C. Ming-Hua, N. Rui-Lin, O. Tanaka, *Phytochemistry* **1995**, 39, 591.
- [21] R.-Q. Sun, Z.-J. Jia, D.-L. Cheng, *Phytochemistry* **1991**, 30, 2707.
- [22] R.-Q. Sun, Z.-J. Jia, *Phytochemistry* **1991**, 30, 3480.
- [23] R.-Q. Sun, J.-C. Chen, *Phytochemistry* **1996**, 44, 505.

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